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Kinetic resolution of hydroxyalkanephosphonates catalyzed by *Candida antarctica* lipase B in organic media[☆]

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Abstract—A series of hydroxyalkanephosphonates were studied as substrates for CALB catalyzed acetylation with emphasis on enantioselectivity and chemical structure of substrates. Some hydroxyalkanephosphonates could be resolved successfully to give both (*R*)- and (*S*)-isomers with high enantiomeric excess. © 2002 Elsevier Science Ltd. All rights reserved.

1. Introduction

Chiral hydroxyalkanephosphonates have received significant attention due to their potential biological activities as well as their ability to mimic their carboxylic counterparts.¹ Moreover, they also serve as useful precursors for a variety of substituted phosphonates.² These compounds can be prepared by TiCl₄ catalyzed opening of homochiral 1,3-dioxane acetals with trimethyl phosphite,³ diastereoselective addition of homochiral phosphorus derivatives to aldehydes,⁴ enantioselective reduction of ketophosphonates,^{1d,5} enantioselective addition of dialkyl phosphites to aldehydes⁶ and even chemical resolutions.⁷ The traditional synthetic route leading to such compounds sometimes has drawbacks such as harsh reaction conditions, expensive reagents and low chemical yield. A convenient procedure for the preparation is therefore needed as an alternative method.

Meanwhile, the utility of lipases for efficient resolution of alcohols and related compounds is of great importance in organic synthesis.⁸ Hamm Schmidt has exploited such hydrolases for enantioselective hydrolysis of a series of 1-acyloxyphosphonates in an organic-buffer biphasic system at pH 7.0 using an autotitrator.⁹

Klibanov's first report¹⁰ on lipase catalyzed stereoselective esterifications and transesterifications in organic solvents prompted much work on the kinetic resolution of racemic secondary alcohols in organic media, which can overcome the limitations of the traditional method, such as instability

of enzymes, time consuming experimental procedures, comparably lower selectivity, etc. Nowadays biocatalysis in non-conventional media is a well-established methodology. To the best of our knowledge, however, few hydroxyalkanephosphonates undergo lipase catalyzed resolution in organic solvents.¹¹ The effect of the phosphonate moiety on enzymic transesterification is virtually unknown.¹²

Continuing our study on the biotransformations of organophosphorus compounds,¹³ and taking into account the scarcity of lipase catalyzed resolution of such molecules in organic solvents, we wish to report the catalytic behavior of *Candida antarctica* lipase B (CALB), a recently widely used immobilized lipase, for the kinetic resolution of hydroxyalkanephosphonates.

2. Results and discussion

The most commonly used method of lipase catalyzed resolution in organic media is transesterification,¹⁴ and our study started with an investigation of CALB catalyzed acylation of diethyl 1-hydroxyethanephosphonate.

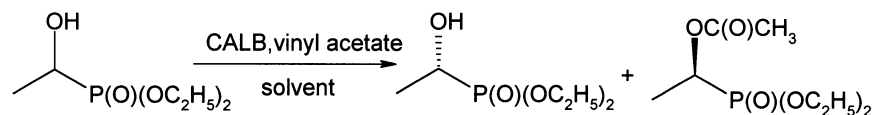
The generally used acylating agents include carboxylic acids, various activated and enol esters, or acyclic aliphatic acid anhydrides, 1-ethoxyvinyl acetate etc.¹⁵ The enzymatic reactions amounted to conversion of one enantiomer of the alcohol into an ester with the other one remaining as the unreacted alcohol. Separation of the ester and unreacted alcohol enabled the resolution of racemic mixtures into their enantiomers. Here we chose vinyl acetate as acyl donor (Scheme 1) since it tautomerizes into acetaldehyde making the reaction irreversible.

In many cases, solvent engineering, i.e. careful selection of the most suitable solvent in the light of physicochemical

[☆] Studies on organophosphorus compounds 115.

Keywords: hydroxyalkanephosphonates; *Candida antarctica* lipase B (CALB); acetylation; optically pure.

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Scheme 1.

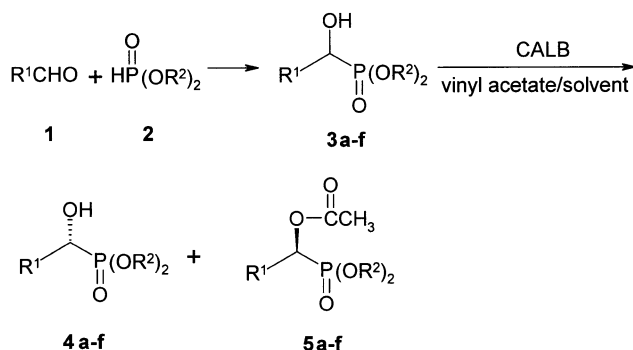
Table 1. Effect of solvent on catalytic acetylation of **1** by CALB

Solvent	E^a	Solvent	E^a
Heptane	6.4	Tetrahydrofuran	>100
Toluene	>100	Vinyl acetate	5.2
Diisopropyl ether	11.31	Acetonitrile	3.1
Benzene	>100	Dioxane	>100

^a The enantiomeric ratio, $E = \ln[(1-c)(1-ees)] / \ln[(1-c)(1+ees)] = \ln[1-c(1+eep)] / \ln[1-c(1-cep)]$, $c = ees / (ees + eep)$.

properties and substrate solubility, can contribute to obtaining the highest enantioselectivity.¹⁶ We investigated the effect of solvent on this enzymatic reaction, and the results are listed in Table 1.

Although the acyl transfer reaction catalyzed by CALB in organic media is believed to follow a bi-bi ping-pong mechanism,¹⁸ and solvent effects have been thoroughly studied in many cases, there still seems no obvious correlation between solvent effect and reaction enantioselectivity.¹⁹ As shown in Table 1, solvents of various types work well here. (The case was also true when several other substrates were tested.) Additionally, the observed rate of reaction differed little with the variation of solvent, and it meant choice of solvent did not represent a serious problem concerning the rate of conversion for practical purposes.



Scheme 2.

Table 2. CALB catalyzed enantioselective acetylation of 1-hydroxyalkane phosphonates

Entry	Substrate	Time (h)	R ¹	R ²	4		5		E
					Yield (%)	ee (%) ^a	Yield (%)	ee (%) ^b	
1	3a	30	Me	Et	43	>95	44	>95	>100
2	3b	30	Me	<i>i</i> -Pr	42	>95	43	>95	>100
3	3c	48	Et	Me	50	50.8	31	>95	>50
4	3d	50	Et	Et	47	79.7	41	>95	>95
5	3e	30	CH ₂ =CH	Et	41	>95	40	>95	>100
6	3f	30	CH ₂ =CH	<i>i</i> -Pr	43	>95	44	>95	>100

^a The ee value was determined by ³¹P NMR using quinine as a solvating agent,²¹ a single peak was considered as an ee value of more than 95%.

^b Ee value was determined by ³¹P NMR after its chemical conversion into alcohols.

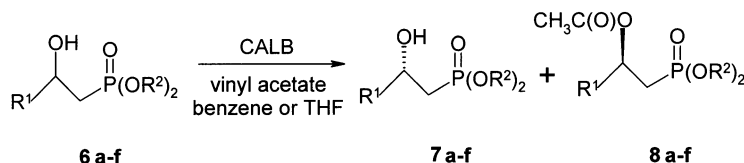
The configuration of the remaining hydroxyethanephosphonate was assigned as (*R*) according to the ³¹P NMR of its ester of (*R*)-Moshers acid.²⁰ Under optimal conditions, (*S*)-substrate was exclusively acetylated to the corresponding acetate to give both optically pure (*R*)-substrate and (*S*)-acetate in nearly 50% yield, respectively. We also carried out this experiment easily on the gram scale.

We extended the substrates to some other 1-hydroxyalkane phosphonates (Scheme 2) which were conveniently prepared from dialkyl phosphite and aldehyde in the presence of a base.⁹ The results were shown in Table 2.

According to the general principle of CALB catalyzed resolution,²² in most cases, the medium group should not be too large. It is demonstrated by us that other groups such as phenyl and propyl are not suitable. In our continuous effort to introduce the trifluoromethyl group to organophosphorus compounds, we wanted to obtain optically pure 1-hydroxy-2,2,2-trifluoroethanephosphonate via this methodology. But there was no trace of the corresponding acetate detected even after one week. Our attempt to resolve dialkyl 2-chloro-1-hydroxyethanephosphonates also failed. The phenomenon that a halogen atom inhibits CALB catalyzed acetylation of hydroxyalkane phosphonates may be ascribed to two reasons. One is that two electron-withdrawing group are not helpful for acetylation. The second may be the non-steric effect.²³

Despite this, CALB shows quite high enantioselectivity in acetylation of (*S*)-hydroxyalkane phosphonates, and this method provides a practical way to prepare optically pure 1-hydroxyalkane phosphonates.

2-Hydroxyalkane phosphonates are also an important class of potential biologically active compounds,^{1d} and they are precursors of 2-aminophosphonates.²⁴ Herein we wish to use CALB catalyzed acetylation for resolution of these compounds (Scheme 3) since few enzymatic resolution routes have been adopted in this field. The results are listed below (Table 3).



Scheme 3.

Table 3. CALB catalyzed enantioselective acetylation of 2-hydroxyalkane phosphonates

Entry	Substrate	Time (h)	R ¹	R ²	7		8		E
					Yield (%)	ee (%)	Yield (%)	ee (%)	
1	6a	30	Me	Me	39	>95	40	>95	>100
2	6b	30	Me	Et	40	>95	39	>95	>100
3	6c	50	Et	Et	43	84.7	35	>95	>100
4	6d	28	Vinyl	Et	40	>95	40	>95	>100
5 ^a	6e	30	ClCH ₂	Et	40	50.2	43	/	<5
6	6f	120	CF ₃	Et	No reaction				

^a The ee value of **7e** is not determined, and the conversion of the reaction is about 50% according to ³¹P NMR of the mixture.

2-Hydroxy-3-chloropropanephosphonate underwent acetylation smoothly because the electron-withdrawing phosphonate moiety is far away from its chiral center, but its enantioselectivity was quite poor due to the non-steric effect. Compound **6f**, however, still resisted the enzymatic reaction due to the strong electron-withdrawing effect of the trifluoromethyl group.

Obviously, methyl, ethyl, and vinyl groups still serve as suitable medium groups here. We observe that 2-hydroxyalkane phosphonates reacted slightly faster than the corresponding 1-hydroxyalkane phosphonates.

That (*S*)-1- and (*R*)-2-hydroxyalkane phosphonates were preferentially acetylated to acetates by CALB is in accordance with the general rule predicted for CALB catalyzed resolutions (Fig. 1).

It is interesting to note that when diethyl 3-hydroxybutane phosphonate, prepared via reduction of diethyl 3-oxobutanephosphonate, was subjected to CALB catalyzed acetylation, it disappeared within 50 h under the same conditions (Scheme 4). The comparably lower enantioselectivity may be due to the fact that the enzyme–substrate binding transition state of both enantiomers differs less than that of 1- and 2-hydroxyalkane phosphonates.

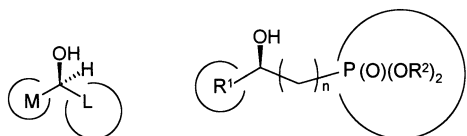


Figure 1. Configuration of the preferential enantiomer of hydroxyalkane phosphonate acetylation catalyzed by CALB.



Scheme 4.

3. Conclusion

In conclusion, some 1-, and 2-hydroxyalkane phosphonates have been successfully resolved by a CALB catalyzed acetylation process. The high enantioselectivity achieved in these reactions as well as the simplicity of the procedure make this strategy a useful alternative for the preparation of optically pure hydroxyalkane phosphonates. The scope and limitations of enzymatic methodology in this field are currently under investigation.

4. Experimental

4.1. General methods

IR spectra were recorded on a Shimadzu IR-440 spectrometer. EI mass spectra (MS) were run on a HP-5989A mass spectrometer. ¹H and ³¹P NMR spectra were recorded on a Bruker AMX-330 (300 MHz) spectrometer in CDCl₃ and chemical shifts were reported in ppm downfield relative to TMS (internal standard) and 80% phosphorus acid (external standard) in phosphorus spectra.

CALB (Novozym 435) is a gift from Novo Nordisk Co. Solvents used for enzymatic reactions were dried by standard methods and stored over 4A sieve before use.

4.2. General procedure for CALB catalyzed acetylation of hydroxyalkane phosphonates

To a stirred solution of hydroxyalkane phosphonate (100 mg) in benzene (3 ml) or THF (3 ml) was added vinyl acetate (0.5 ml). The reaction was started by addition of CALB (100 mg). The mixture was maintained at 45°C. When the reaction proceeded to certain conversion, the enzyme was filtered, washed with 3 ml acetone. The volatile solvent was removed under reduced pressure and the residue was subjected to flash chromatography to furnish almost pure hydroxyalkane phosphonates and their acetates.

4.2.1. (R)-Diethyl 1-hydroxyethanephosphonate (4a).²⁵ Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = -5.5$ (c 0.5, CH₃OH); IR (neat) 3317, 2984, 1220, 1053, 1029 cm⁻¹; ¹H NMR (δ): 4.10–4.17 (4H, m, P(O)(OCH₂CH₃)₂), 4.00 (1H, dq, *J*=3.5, 7.1 Hz, *CHOH*), 3.98 (1H, br, s *OH*), 1.40 (3H, dd, *J*=7.1, 17.5 Hz, CH₃CH(OH)), 1.28–1.33 (6H, m, P(O)(OCH₂CH₃)₂); EIMS (*m/z*): 183 (M+1), 165, 155, 43, 138, 111, 82, 45.

4.2.2. (S)-Diethyl 1-acetyloxyethanephosphonate (5a). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +26.5$ (c 2.0, CHCl₃); IR (neat) 2933, 1740, 1237, 1049, 1027 cm⁻¹; ¹H NMR (δ): 5.27 (1H, dq, *J*=7.2, 8.4 Hz, *CHP*), 4.12–4.23 (4H, m, P(O)(OCH₂CH₃)₂), 2.12 (3H, s, CH₃CO), 1.49 (3H, dd, *J*=7.1, 16.9 Hz, CH₃CHP), 1.31–1.37 (6H, m, P(O)(OCH₂CH₃)₂); EIMS (*m/z*): 225 (M+1), 180, 152, 138, 111, 82, 43; Anal. Calcd for C₈H₁₇O₅P: C, 42.86; H, 7.64. Found: C, 42.61; H, 7.41.

4.2.3. (R)-Diisopropyl 1-hydroxyethanephosphonate (4b).⁹ Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = -6.1$ (c 1.2, CHCl₃); IR (neat) 3317, 2980, 1385, 1220, 1044, 1023 cm⁻¹; ¹H NMR (δ): 4.70–4.79 (2H, m, P(OCHMe₂)₂), 4.38 (1H, br, s, *OH*), 3.97 (1H, dq, *J*=3.1, 6.9 Hz, CH₃CH(OH)), 1.42 (3H, dd, *J*=7.1, 17.2 Hz, CH₃CH(OH)), 1.33 (12H, overlapping, P(OCHMe₂)₂); EIMS (*m/z*): 211 (M+1), 193, 166, 151, 109, 43.

4.2.4. (S)-Diisopropyl 1-acetyloxyethanephosphonate (5b). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +20.10$ (c 1.7, CHCl₃); IR (neat) 2983, 1752, 1229, 1107, 1044, 1028 cm⁻¹; ¹H NMR (δ): 5.18 (1H, dq, *J*=7.2, 8.5 Hz), 4.76 (2H, m, P(OCHMe₂)₂), 2.10 (3H, s, CH₃CO), 1.44 (3H, dd, *J*=7.0, 8.9 Hz, CH₃CH(OH)), 1.30 (12H, dd, *J*=5.9, 11.7 Hz, P(OCHMe₂)₂); EIMS (*m/z*): 253 (M+1), 211, 166, 151, 124, 43; Anal. Calcd for C₁₀H₂₁O₅P: C, 47.62; H, 8.39. Found: C, 47.61; H, 8.34.

4.2.5. (R)-Dimethyl 1-hydroxypropanephosphonate (4c).⁹ Colorless oil, 50.8% ee, $[\alpha]_{\text{D}}^{25} = -4.3$ (c 0.5, CHCl₃); IR (neat) 3317, 2962, 1217, 1033 cm⁻¹; ¹H NMR (δ): 3.79–3.88 (2H, OH, *CHP*), 3.87 (6H, d, *J*=10.3 Hz, P(OCH₃)₂), 1.73–1.84 (2H, m, CH₃CH₂CH), 1.10 (3H, t, *J*=7.4 Hz, CH₃CH₂CH); EIMS (*m/z*): 169 (M+1), 58, 43.

4.2.6. (S)-Dimethyl 1-acetyloxypropanephosphonate (5c). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +44.2$ (c 0.25, CHCl₃); IR (neat) 2962, 1751, 1228, 1032 cm⁻¹; ¹H NMR (δ): 5.14–5.21 (1H, m, *CHP*), 3.75 (6H, 2d, *J*=9.3 Hz, P(OCH₃)₂), 2.13 (3H, s, CH₃CO), 1.85 (2H, m, CH₃CH₂CHP), 0.98 (3H, t, *J*=7.2 Hz, CH₃CH₂CHP); EIMS (*m/z*): 169, 124, 110, 79, 43; Anal. Calcd for C₇H₁₅O₅P: C, 40.01; H, 7.19. Found: C, 40.01; H, 7.19.

4.2.7. (R)-Diethyl 1-hydroxypropanephosphonate (4d).^{5a} Colorless oil, 79.7% ee, $[\alpha]_{\text{D}}^{25} = -4.9$ (c 1.5, CHCl₃); IR (neat) 3312, 2982, 1213, 1048, 1027 cm⁻¹; ¹H NMR (δ): 4.12–4.21 (4H, m, P(OCH₂CH₃)₂), 3.89 (1H, br, s, *OH*), 3.74–3.80 (1H, m, CH₃CH₂CH(OH)), 1.77–1.81 (2H, m, CH₃CH₂CH), 1.31–1.36 (6H, m, P(O)(OCH₂CH₃)₂), 1.07 (3H, t, *J*=7.4 Hz, CH₃CH₂CH); EIMS (*m/z*): 197 (M+1), 179, 138, 111, 82, 65.

4.2.8. (S)-Diethyl 1-acetyloxypropanephosphonate (5d). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +41.3$ (c 1.0, CHCl₃); IR (neat) 2981, 1751, 1327, 1228, 1023; ¹H NMR (δ): 5.07–5.15 (1H, m, *CHP*), 4.01–4.12 (4H, m, P(OCH₂CH₃)₂), 2.06 (3H, s, CH₃CO), 1.71–1.87 (2H, m, CH₃CH₂CHP), 1.18–1.28 (6H, m, P(O)(OCH₂CH₃)₂), 0.95 (3H, t, *J*=7.4 Hz, CH₃CH₂CHP); EIMS (*m/z*): 239 (M+1), 180, 152, 138, 111, 82, 43; Anal. Calcd for C₉H₁₉O₅P: C, 45.38; H, 8.04. Found: C, 45.11; H, 7.82.

4.2.9. (R)-Diethyl 1-hydroxy-2-propenylphosphonate (4e).²⁶ Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = -12.7$ (c 0.8, CHCl₃); IR (neat): 3293, 2986, 1641, 1238, 1201, 1052, 1028 cm⁻¹; ¹H NMR (δ): 5.27–6.03 (3H, H of Vinyl), 4.73 (1H, br, s, *OH*), 4.35 (1H, dd, *J*=5.4, 13.7 Hz, *CHP*), 4.13–4.23 (m, 4H, P(OCH₂CH₃)₂), 1.33 (6H, m, P(OCH₂CH₃)₂); EIMS (*m/z*): 195 (M+1), 177, 149, 151, 111, 82, 57.

4.2.10. (S)-Diethyl 1-acetyloxy-2-propenylphosphonate (5e). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +23.8$ (c 0.9, CHCl₃); IR (neat): 2987, 1753, 1643, 1262, 1226, 1052, 1025 cm⁻¹; ¹H NMR (δ): 5.25–5.90 (4H, H of Vinyl), 5.85 (1H, overlapping, *CHP*), 4.07–4.12 (4H, m, P(OCH₂CH₃)₂), 2.11 (3H, s, CH₃CO), 1.23–1.28 (6H, m, P(OCH₂CH₃)₂); EIMS (*m/z*): 194, 177, 165, 138, 111, 82, 43; Anal. Calcd for C₉H₁₇O₅P: C, 45.77; H, 7.25. Found: C, 45.79; H, 7.36.

4.2.11. (R)-Diisopropyl 1-hydroxy-2-propenylphosphonate (4f).²⁷ Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = -14.4$ (c 0.7, CHCl₃); IR (neat) 3297, 2970, 1645, 1240 cm⁻¹; ¹H NMR (δ): 5.27–6.01 (3H, H of Vinyl), 4.71–4.79 (2H, m, P(OCHMe₂)₂), 4.43 (1H, dd, *J*=5.4, 13.3 Hz, *CHP*), 3.96 (1H, OH), 1.26–1.44 (m, 12H, overlapping, P(OCHMe₂)₂); EIMS (*m/z*): 223 (M+1), 193, 181, 165, 139, 109, 43.

4.2.12. (S)-Diisopropyl 1-acetyloxy-2-propenylphosphonate (5f). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +26.5$ (c 1.3, CHCl₃); IR: 2984, 1753, 1634, 1261, 1226, 999 cm⁻¹; ¹H NMR (δ): 5.25–5.90 (4H, H or Vinyl; *CHP*), 4.76 (2H, m, P(OCHMe₂)₂), 3.96 (1H, dd, *J*=5.4, 13.5 Hz), 2.17 (3H, s, CH₃CO), 1.33 (12H, dd, *J*=6.3, 7.9 Hz) P(OCHMe₂)₂); EIMS (*m/z*): 265 (M+1), 193, 163, 138, 121, 43; Anal. Calcd for C₁₁H₂₁O₅P: C, 50.00; H, 8.01. Found: C, 50.06; H, 7.96.

4.2.13. (S)-Dimethyl 2-hydroxypropanephosphonate (7a).^{1d} Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +13.6$ (c 1.1, CHCl₃); IR (neat) 3402, 2962, 1227, 1033 cm⁻¹; ¹H NMR (δ): 4.06–4.21 (1H, m, CH₃CHOH), 3.72 (6H, 2d, *J*=10.9 Hz, P(OCH₃)₂), 3.34 (1H, s, *OH*), 1.92–2.00 (2H, dd, *J*=6.3, 17.8 Hz, CH₂P), 1.19–1.32 (3H, m, CH₃CHOH); EIMS (*m/z*): 169 (M+1), 151, 124, 109, 94, 79.

4.2.14. (R)-Dimethyl 2-acetyloxypropanephosphonate (8a). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +5.7$ (c 1.0, CHCl₃); IR (neat): 2960, 1739, 1245, 1033 cm⁻¹; ¹H NMR (δ): 5.08–5.11 (1H, m, CH₃CHCH₂P), 3.68 (6H, d, *J*=10.9 Hz, P(OCH₃)₂), 2.1 (3H, s, CH₃CO), 1.90–2.15 (2H, m, CH₂P), 1.31 (3H, d, *J*=6.4 Hz, CH₃CHCH₂P); EIMS (*m/z*): 167, 151, 135, 124, 109, 79, 43; Anal. Calcd for C₇H₁₅O₅P: C, 40.00; H, 7.19. Found: C, 39.97; H, 7.16.

4.2.15. (S)-Diethyl 2-hydroxypropanephosphonate (7b).^{1d} Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +15.2$ (c 0.25, CHCl_3); IR: 3399, 2983, 1234, 1048 cm^{-1} ; $^1\text{H NMR}$ (δ): 4.08–4.23 (5H, m, CH_3CHOH ; $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 3.66 (1H, br, s, OH), 1.96 (2H, dd, $J=6.3$, 17.1 Hz, CH_2P), 1.36–1.38 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 1.31 (3H, dd, $J=2.4$, 5.9 Hz, CH_3CH); EIMS (m/z): 197 (M+1), 179.

4.2.16. (R)-Diethyl 2-acetyloxypropanephosphonate (8b). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +9.7$ (c 0.5, CHCl_3); IR: 2986, 1740, 1244, 1027, 958 cm^{-1} ; $^1\text{H NMR}$ (δ): 5.13–5.18 (1H, m, $\text{CH}_3\text{CHCH}_2\text{P}$), 4.03–4.13 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 1.81–2.42 (2H, m, CH_2P), 2.08 (3H, s, CH_3CO), 1.31 (2H, dd, $J=3.0$, 10.2 Hz, $\text{CH}_3\text{CHCH}_2\text{P}$), 1.22–1.28 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$); EIMS (m/z): 239 (M+1), 195, 179, 151, 43; Anal. Calcd for $\text{C}_9\text{H}_{19}\text{O}_5\text{P}$: C, 45.38; H, 8.04. Found: C, 45.20; H, 7.84.

4.2.17. (S)-Diethyl 2-hydroxybutanephosphonate (7c).²⁸ Colorless oil, 84.7% ee, $[\alpha]_{\text{D}}^{25} = +13.1$ (c 0.21, CHCl_3); IR (neat) 3394, 2982, 1225, 1056, 1029, 967 cm^{-1} ; $^1\text{H NMR}$ (δ): 4.11–4.15 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 3.83 (2H, m, CHOH), 1.86–2.03 (2H, m, CH_2P), 1.56–1.58 (2H, m, $\text{CH}_3\text{CH}_2\text{CHOH}$), 1.31–1.37 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 0.95–0.99 (3H, m, $\text{CH}_3\text{CH}_2\text{CHOH}$); EIMS (m/z): 211 (M+1), 193, 181, 153, 125.

4.2.18. (R)-Diethyl 2-acetyloxybutanephosphonate (8c). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +8.2$ (c 0.7, CHCl_3); IR (neat) 2980, 1740, 1243, 1055, 1026, 967 cm^{-1} ; $^1\text{H NMR}$ (δ): 5.04 (1H, CHCH_2P), 3.99–4.10 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 2.10 (3H, s, CH_3CO), 1.81–2.12 (2H, CH_2P), 1.58–2.08 (2H, m, $\text{CH}_3\text{CH}_2\text{CHCH}_2\text{P}$), 1.24–1.29 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 0.98 (3H, t, $J=7.1$ Hz, $\text{CH}_3\text{CH}_2\text{CHCH}_2\text{P}$); EIMS (m/z): 253 (M+1), 211, 193, 43; Anal. Calcd for $\text{C}_{10}\text{H}_{21}\text{O}_5\text{P}$: C, 47.62; H, 8.39. Found: C, 47.49; H, 8.39.

4.2.19. (S)-Diethyl 2-hydroxy-3-butenylphosphonate (7d). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +5.6$ (c 5.6, CHCl_3); IR (neat) 3364, 2985, 1630, 1224, 1058, 1029 cm^{-1} ; $^1\text{H NMR}$ (δ): 5.12–5.96 (3H, H of Vinyl), 4.49–4.56 (1H, m, CHOH), 4.06–4.19 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 3.58 (1H, br, s, OH), 2.03 (2H, dd, $J=6.5$, 17.2 Hz, CH_2P), 1.32–1.36 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$); EIMS (m/z): 208 (M), 191, 179, 152, 125, 97; Anal. Calcd for $\text{C}_8\text{H}_{17}\text{O}_4\text{P}$: C, 46.15; H, 8.23. Found: C, 46.15; H, 8.10.

4.2.20. (R)-Diethyl 2-acetyloxy-3-butenylphosphonate (8d). Colorless oil, >95% ee, $[\alpha]_{\text{D}}^{25} = +2.5$ (c 1.4, CHCl_3); IR (neat) 2987, 1743, 1647, 1240, 1025 cm^{-1} ; $^1\text{H NMR}$ (δ): 5.17–5.89 (4H, H of Vinyl; CHCH_2P), 4.02–4.11 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 2.1 (3H, s, CH_3CO), 2.00–2.20 (2H, m, CH_2P), 1.22–1.31 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$); EIMS (m/z): 251 (M+1), 191, 163, 43; Anal. Calcd for $\text{C}_{10}\text{H}_{19}\text{O}_5\text{P}$: C, 48.00; H, 7.65. Found: C, 48.19; H, 7.61.

4.2.21. (R)-Diethyl 3-chloro-2-hydroxypropanephosphonate (7f).²⁸ Colorless oil, 50.2% ee, $[\alpha]_{\text{D}}^{25} = +3.5$ (c 2.2, CHCl_3); IR (neat) 3384, 2965, 1220, 1025 cm^{-1} ; $^1\text{H NMR}$ (δ): 4.14–4.31 (5H, m, CHOH ; $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 3.85 (1H, br, s, OH), 3.61 (2H, d, $J=5.2$ Hz, ClCH_2), 2.04–2.17 (2H, m, CH_2P), 1.25–1.36 (6H, m,

$\text{P}(\text{OCH}_2\text{CH}_3)_2$); EIMS (m/z): 231 (M+1), 181, 139, 125, 111, 93.

4.2.22. (S)-Diethyl 3-chloro-2-acetyloxypropanephosphonate (8f). Colorless oil, ee value not determined; IR (neat) 2986, 1747, 1233, 1028, 969 cm^{-1} ; $^1\text{H NMR}$ (δ): 5.30 (1H, m, ClCH_2CH), 4.05–4.16 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 3.67–3.80 (2H, m, ClCH_2), 2.11–2.27 (2H, m, CH_2P), 2.10 (3H, s, CH_3CO), 1.29–1.34 (3H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$); EIMS (m/z): 273 (M), 213, 195, 181, 125, 43; Anal. Calcd for $\text{C}_9\text{H}_{18}\text{O}_5\text{PCl}$: C, 39.65; H, 6.65. Found: C, 39.88; H, 6.67.

4.2.23. Diethyl 3-hydroxybutanephosphonate (9). Colorless oil, IR (neat) 3397, 1270, 1033 cm^{-1} ; $^1\text{H NMR}$ (δ): 4.00–4.10 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 3.89 (1H, m, CHOH), 2.90 (1H, br, s, OH), 1.67–1.87 (4H, $\text{CH}_2\text{CH}_2\text{P}$), 1.26–1.32 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 1.16 (3H, d, $J=6.2$ Hz, CH_3CHOH); EIMS (m/z): 195, 137, 121, 111, 82, 55; Anal. Calcd for $\text{C}_8\text{H}_{19}\text{O}_4\text{P}$: C, 45.71; H, 9.11. Found: C, 45.30; H, 8.89.

4.2.24. Diethyl 3-acetyloxybutanephosphonate (10). Colorless oil, IR (neat) 2984, 1737, 1375, 1246, 1057, 1026 cm^{-1} ; $^1\text{H NMR}$ (δ): 4.63 (1H, m, $\text{CH}_3\text{CHCH}_2\text{CH}_2\text{P}$), 4.03–4.13 (4H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 2.02 (3H, s, CH_3CO), 1.73–1.85 (4H, $\text{CH}_2\text{CH}_2\text{P}$), 1.29–1.33 (6H, m, $\text{P}(\text{OCH}_2\text{CH}_3)_2$), 1.21 (3H, d, $J=6.0$ Hz, $\text{CH}_3\text{CHCH}_2\text{CH}_2\text{P}$); EIMS (m/z): 209, 193, 43; Anal. Calcd for $\text{C}_{10}\text{H}_{21}\text{O}_5\text{P}$: C, 47.62; H, 8.39. Found: C, 47.39; H, 8.38.

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